

A new bioanalytical tool for neurotransmitter monitoring in central nervous system diseases

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Contextualization

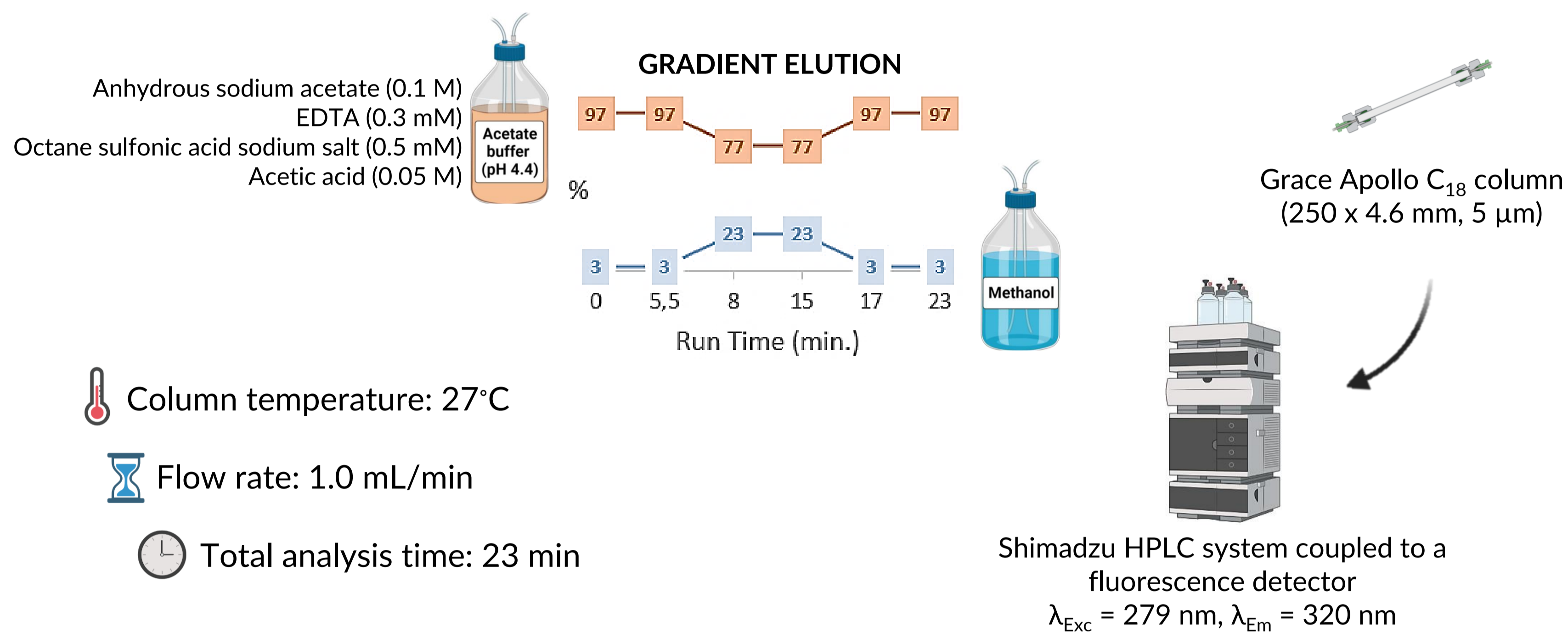
- **Noradrenaline (NA), adrenaline (AD), dopamine (DA) and serotonin (5-HT)** are essential neurotransmitters for normal neuronal function¹;
- Neurotransmitter homeostasis is compromised by aging and neurological diseases such as **Alzheimer's disease** and **depression**^{2,3};
- The dynamic interaction between the monoamine pathways sustains their simultaneous monitorization^{3,4,5};
- Monitoring the levels of neurotransmitters, their metabolites, namely DA metabolites 3,4-dihydroxyphenylacetic acid (DOPAC) and homovanillic acid (HVA), and their precursors, such 5-HT precursor tryptophan (TRYP), in biological matrices presents high relevance particularly to assess disease severity and progression¹.

Objectives

- **To develop** an accurate and precise High-Performance Liquid Chromatography (HPLC) method to quantify NA, AD, DA, DOPAC, HVA, TRYP and 5-HT in mouse brain tissues;
- **To validate** the HPLC method according to the ICH guideline M10 on Bioanalytical Method Validation and study sample analysis;
- **To demonstrate its application** in preclinical *in vivo* studies.

Method development

➤ Chromatography conditions



➤ Tissue preparation

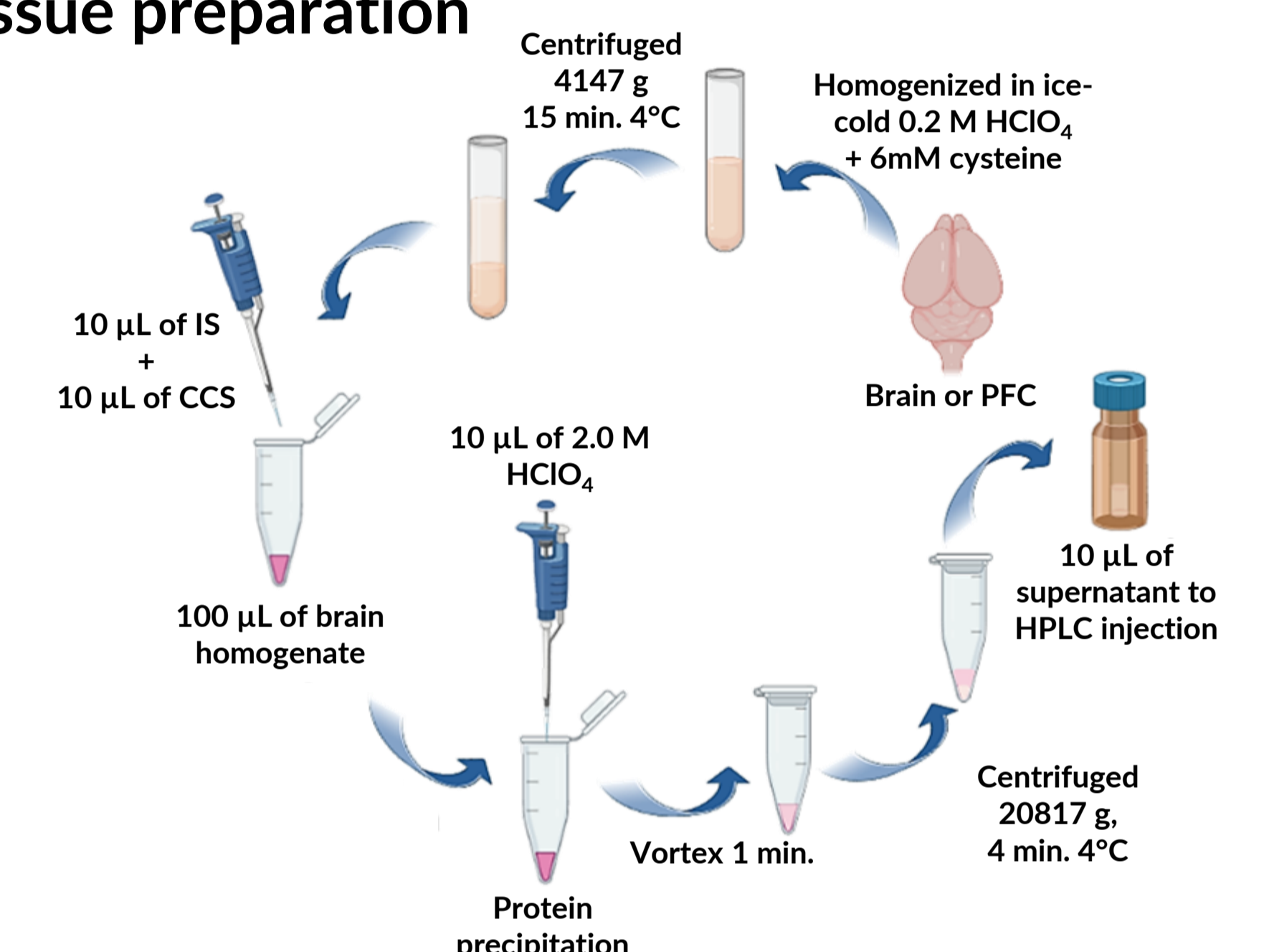


Figure 1. Brain tissue preparation by protein precipitation. CCS: combined calibration standard; IS: internal standard; PFC: prefrontal cortex.

Method validation

➤ ICH guideline requirements for method validation⁶:

- Selectivity
- Specificity
- Carry-over
- Calibration curve range and linearity
- Accuracy, precision and recovery
- Stability

➤ 1 NA, AD, DOPAC, DA, HVA, 5-HT and TRYP → Analytes that are also Endogenous Compounds

Background Subtraction Approach (ICH guideline for bioanalytical method validation)⁶

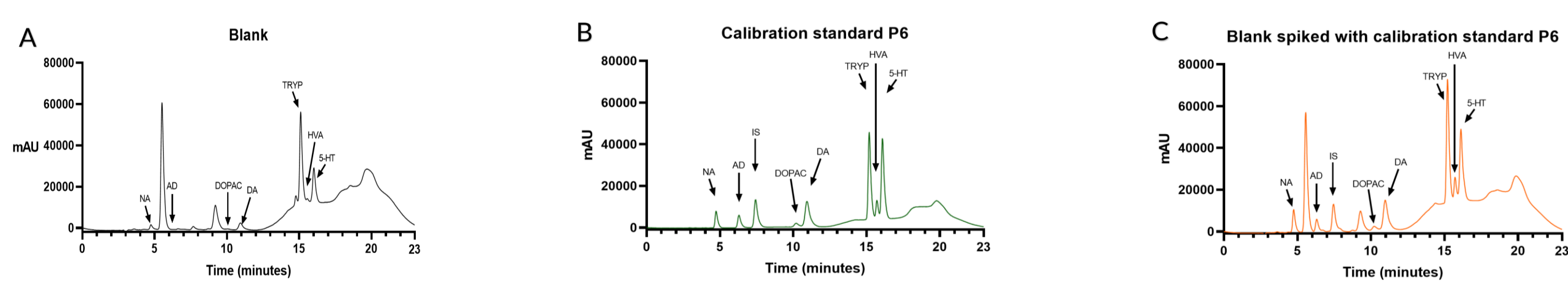


Figure 2. Chromatogram representative of a blank sample of mice brain homogenate (A), the calibration standard P6 (B) and the blank spiked with the calibration standard P6 (C).

5-HT: serotonin; AD: adrenaline; DA: dopamine; DOPAC: 3,4-dihydroxyphenylacetic acid; HVA: homovanillic acid; NA: noradrenaline; TRYP: tryptophan

➤ 2 No interference of:

- Norfloxacin
 - Ofloxacin
 - Prulifloxacin
 - Gatifloxacin
 - Pazufloxacin
 - Amitriptyline
 - Antipyrine
 - Pitavastatin
 - Atorvastatin
 - Metformin
 - Clinofloxacin
 - Lacosamide
 - Levetiracetam
 - Zonisamide
 - Perampnel
 - Cannabidiol
 - Cannabigerol
 - Cannabichromene
 - Cannabinol
 - Cannabidiol
- 3 Not observed

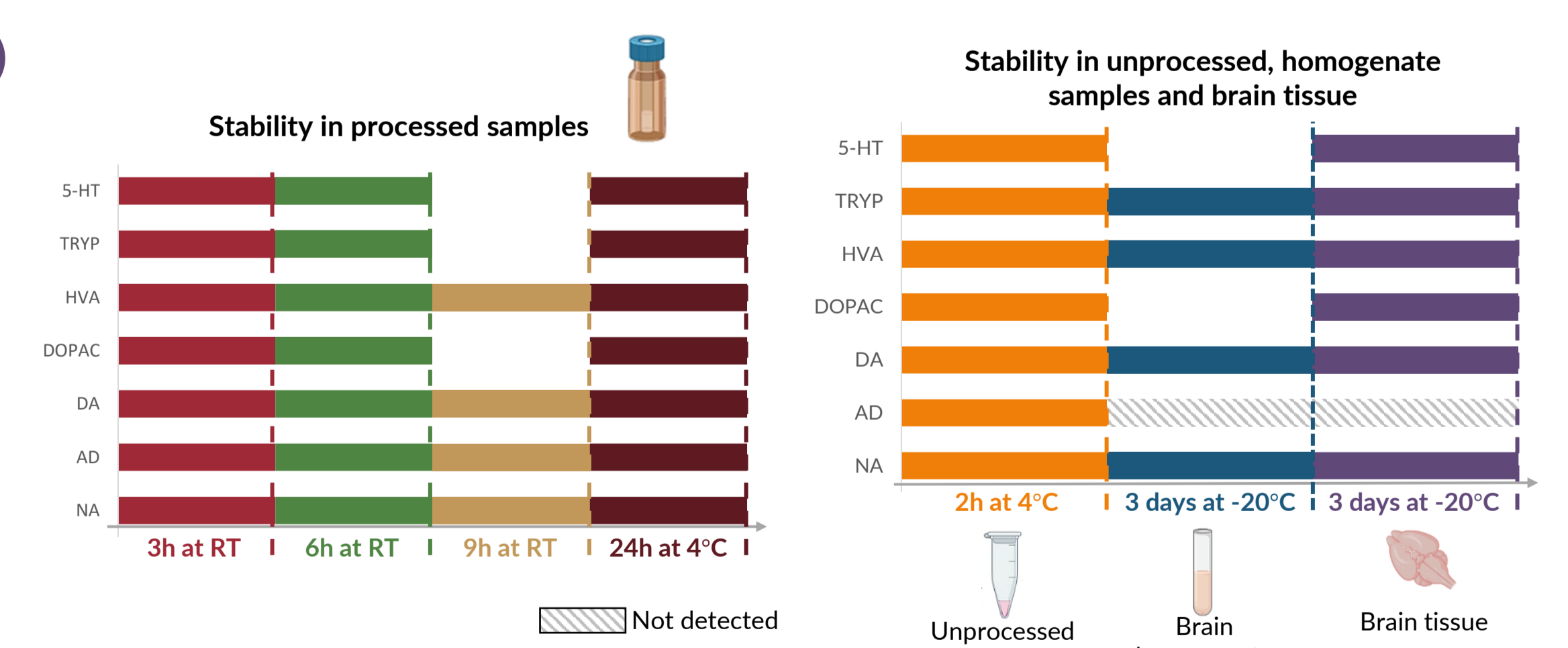
4 Table 1. Validation parameters of the analytes.

Analyte	Calibration curves range (ng/mL)	Linearity (R-squared ²)	Accuracy (%)	Precision (CV)	Recovery (%)
NA	10-400	0.997	-8.72 - 1.56	1.37 - 11.39	101.80 ± 11.39
AD	10-200	0.988	-7.62 - 14.71	1.01 - 7.98	70.88 ± 6.17
DOPAC	50-300	0.998	-9.97 - 5.90	4.36 - 15.29*	67.73 ± 12.84
DA	10-800	0.997	-7.60 - 18.87*	1.19 - 16.22*	89.52 ± 11.69
HVA	55-300	0.997	-8.69 - 5.58	0.95 - 3.81	84.53 ± 8.20
TRYP	35-1000	0.997	-12.73 - 6.03	-12.73 - 14.56	80.86 ± 11.59
5-HT	35-300	0.999	-2.60 - 14.34	1.29 - 8.56	86.95 ± 8.48

Representation of the maximum ranges and values in inter- and intra-day validation (n=5); Accuracy (%) = 100 × [(Measured concentration of spiked sample - endogenous concentration) - Nominal concentration] / Nominal concentration; CV, coefficient of variation (%) = 100 × (Standard Deviation/Mean); Recovery (%) = Mean ± Standard Deviation.

*Values obtained at the lower limit of quantification (LLOQ) quality sample.

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Method application

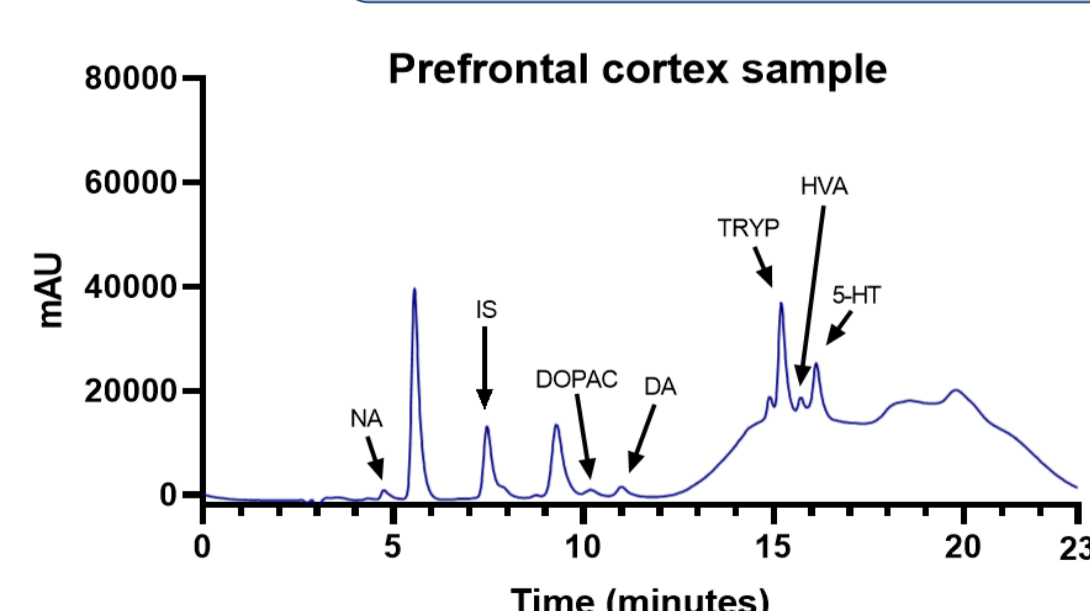


Figure 3. Chromatogram of a prefrontal cortex sample collected from mice treated with 55 μmol/Kg of ketamine intraperitoneally.

5-HT: serotonin; DA: dopamine; DOPAC: 3,4-dihydroxyphenylacetic acid; HVA: homovanillic acid; IS: Internal Standard; NA: noradrenaline; TRYP: tryptophan.

- Applied to study the chronic antidepressant effect of sertraline (DOI:10.1016/j.ejpb.2023.12.002)
- Applied in a genetic mouse model of autism spectrum disorders
- Applied to study the acute antidepressant effect of ketamine

Final remarks

- The method was successfully developed and validated according to the ICH M10 guideline for bioanalytical method validation;
- It resorts to native fluorescence of the analytes, avoiding unspecific detections;
- It enables a sustainable sample preparation, saving time and resources;
- It allows simultaneous quantification of 7 compounds (4 neurotransmitters, 2 metabolites and 1 precursor) in mouse brain and prefrontal cortex homogenates;
- It has demonstrated reproducible application in *in vivo* preclinical studies.
- It can be further applied in several animal models of neurological diseases.

References:

- [1] G. E. De Benedetto et al., (2014). J. Pharm. Biomed. Anal., 98, 266-270. [2] M. Mather, (2021). Semin. Cell Dev. Biol., 116(May), 108-124. [3] C. Kraus, et al., (2017). Neurosci. Biobehav. Rev., 77, 317-326. [4] Correia et al., Int. J. Mol. Sci., 2022. [5] Kanova et al., Int. J. Mol. Sci., 2021. [6] International Council Harmonised Guideline (2019). Bioanalytical Method Validation M10.



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